

- (17) Robin, M. B. "Higher Excited States of Polyatomic Molecules"; Academic Press: New York, 1975; Vol. II, pp 68-75.
 (18) Moore, W. R.; Andersen, H. W.; Clark, S. D.; Ozretich, T. M. *J. Am. Chem. Soc.* **1971**, *93*, 4932.
 (19) The authors acknowledge the financial support of this research by the National Science Foundation and the National Institutes of Health.
 (20) On sabbatical leave (1977-1978) from Otterbein College, Westerville, Ohio.

Jerry A. Jenkins,²⁰ Robert E. Doehner, Jr.
 Leo A. Paquette*

Evans Chemical Laboratories, The Ohio State University
 Columbus, Ohio 43210

Received October 1, 1979

Ficisterol
(23-Ethyl-24-methyl-27-norcholesta-5,25-dien-3 β -ol).
A Biosynthetically Unprecedented Sterol from the
Marine Sponge *Petrosia ficiformis*¹

Sir:

Petrosterol (**1**, 26,27-cycloaplysterol), a novel cyclopropane-containing sterol, has recently been identified in a Pacific sponge, *Halichondria* sp.² and as the major sterol from *Petrosia ficiformis* collected in the Bay of Naples.^{3,4} In view of our interest in the biosynthetic origin of the cyclopropane function in the sterol side chain, we have carefully analyzed the minor and trace components of *P. ficiformis* with the hope of identifying possible related biosynthetic intermediates. We now report the presence of a new C₂₉ sterol with an unprecedented side chain, shown to be 23-ethyl-24-methyl-27-norcholesta-5,25-dien-3 β -ol (**2**), which we have named ficisterol.

Argent TLC separation of *P. ficiformis* sterol acetates³ using 5:2⁶ (v/v) hexane-benzene or hexane-toluene gave several bands, the most polar of which contained four major components. Repeated TLC with hexane-toluene (1:1) and reversed-phase HPLC on C₁₈ μ Bondapak⁷ gave **2** (0.6% of total sterol fraction), M⁺ 412.3701 (C₂₉H₄₈O), whose GC (OV-17 column) retention time (cholesterol, 1.00) was 1.58, acetate mp 99-100 °C. Since the presence of the usual Δ^5 -3 β -hydroxy sterol nucleus was demonstrated⁸ by the peaks at *m/z* 271, 255, 231, and 213, the sterol must possess one degree of unsaturation in the side chain. The 360-MHz ¹H NMR spectrum in C₆D₆ (δ , parts per million) of the acetate of **2** shows six methyl group signals (see Table I), with one of the signals clearly a triplet, suggesting the presence of a -CH₂-CH₃ entity on the side chain. Significantly, doublets due to a terminal isopropyl group are absent. A multiplet at 5.85 is assigned to the vinylic proton at C-25, while other easily recognizable signals are 5.38 (C-6 H) and 4.85 (m) (3 α -H acetate). A pair of doublets (2 H) at 5.03 and 5.06 is typical for a terminal methylene group,⁹ and double resonance experiments showed that these protons at C-26 are coupled with the C-25 vinylic hydrogen at 5.85.

Irradiation of the allylic C-24 proton at 2.27 collapsed the 28-methyl doublet at 0.95 to a singlet, and at the same time simplified the complex vinylic proton region at 5.85. Irradiation of the C-26 geminal protons also simplified the vinylic C-25 hydrogen and the resonance due to the C-24 allylic proton. Irradiation of the vinylic C-25 hydrogen collapses the geminal C-26 protons and simplified the allylic C-24 hydrogen. The presence of part of the side chain as a 3-substituted but-1-ene is thus established, which with the addition of the requisite ethyl group leads to structure **2** or **4**, both of them being unprecedented in terms of a 22- or 23-ethyl substituent.

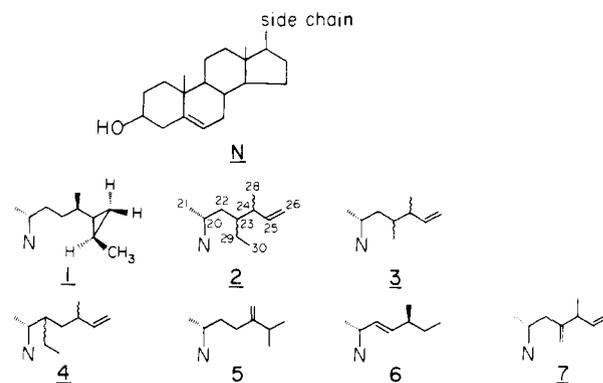
Triple irradiation experiments performed by simultaneously irradiating the 28-methyl group and C-25 vinylic hydrogen clearly produced a doublet (*J* ~ 5 Hz) for the allylic hydrogen

Table I. ¹H Chemical Shifts of the Acetates of Ficisterol (**2**) and 23,24-Dimethyl-27-norcholesta-5,25-dien-3 β -ol (**3**) (360 MHz, C₆D₆)

carbon	2	3
18-CH ₃	0.686 (s)	0.684 (s)
19-CH ₃	0.918 (s)	0.925 (s)
21-CH ₃	0.984 (d, <i>J</i> = 6.5) ^a	1.017 (d, <i>J</i> = 6.9)
30-CH ₃	0.887 (t, <i>J</i> = 7.3)	
29-CH ₃		0.867 (d, <i>J</i> = 6.7)
28-CH ₃	0.955 (d, <i>J</i> = 6.9)	0.972 (d, <i>J</i> = 6.4)
C-25 H	5.85 (m)	5.78 (m)
C-26 (2 H)	5.03, 5.06 (d)	5.03, 5.06 (d)
C-6 H	5.35 (m)	5.38 (m)
C-3 (COCH ₃)	1.75 (s)	1.75 (s)
3- α -H of acetate	4.85 (m)	4.86 (m)

^a Coupling constants *J* are in hertz.

at C-24, thus demonstrating that position 23 is monosubstituted (i.e., **2**). The lower homologue **3** (C₂₈H₄₆O, M⁺ 398) has also been isolated as a 1:1 acetate mixture with 24-methylenecholesterol (**5**), but the difference in the chemical shift of the C-21 and C-28 methyl protons (Table I) of the acetate suggest differences in stereochemistry.



Ficisterol (**2**) is biosynthetically intriguing since it lacks the normal terminal isopropyl functionality and therefore resembles ocellasterol (**6**), isolated from a marine annelid.¹⁰ Even more unusual is the presence of the C-23 ethyl substituent; although several marine sterols (e.g., gorgosterol,¹¹ 23,24-dimethylcholesta-5,22-dien-3 β -ol,¹² and dinosterol¹³) have been shown to be methylated at C-23 and C-24, ficisterol (**2**) is the first example of C-23 ethylation in a naturally occurring sterol. This suggests that methylation of a hitherto undetected 23-methylene sterol (e.g., **7**) is a new mode of sterol-side-chain bioalkylation and we are actively engaged in searching in marine organisms for further examples.

Acknowledgments. We acknowledge financial support from the National Institutes of Health (Grants GM-06840 and AM-04257) and use of a 360-MHz NMR spectrometer made possible by grants from the National Science Foundation (GP-23633) and the NIH (RR-00711). We thank Annemarie Wegmann for mass spectral determinations. M.W.K. was the recipient of a NATO research grant, SRG-10.

References and Notes

- (1) Part 15 of the Stanford series, "Minor and Trace Sterols in Marine Invertebrates". For preceding paper, see Delseth, C.; Kashman, Y.; Djerassi, C. *Helv. Chim. Acta* **1979**, *62*, 2037-2045.
- (2) Ravi, B.; Kokke, W. C. M. C.; Delseth, C.; Djerassi, C. *Tetrahedron Lett.* **1978**, 4379-4380.
- (3) Sica, D.; Zollo, F. *Tetrahedron Lett.* **1978**, 837-838.
- (4) Mattia, C. A.; Mazzarella, L.; Puliti, R.; Sica, D.; Zollo, F. *Tetrahedron Lett.* **1978**, 3953-3954.
- (5) Djerassi, C.; Theobald, N.; Kokke, W. C. M. C.; Pak, C. S.; Carlson, R. M. K. *Pure Appl. Chem.* **1979**, *51*, 1815-1828.
- (6) Idler, D. R.; Safe, L. M. *Steroids* **1972**, *19*, 315-324.
- (7) Popov, S.; Carlson, R. M. K.; Wegmann, A.; Djerassi, C. *Steroids* **1976**, *28*, 699-732.

- (8) Zaretskii, Z. V. "Mass Spectrometry of Steroids"; Israel Universities Press: Jerusalem, 1976.
 (9) Jackman, L. M.; Sternhell, S. "Applications of Nuclear Magnetic Resonance Spectroscopy in Organic Chemistry", 2nd ed.; Pergamon Press: New York, 1969; pp 184-192.
 (10) Kobayashi, M.; Mitsunashi, H. *Steroids* **1974**, *24*, 399-410.
 (11) Ling, N. C.; Hale, R. L.; Djerassi, C. *J. Am. Chem. Soc.* **1970**, *92*, 5281-5282.
 (12) Kanazawa, A.; Teshima, S.; Ando, T.; Tomita, S. *Nippon Suisan Gakkaishi* **1974**, *40*, 729.
 (13) Shimizu, Y.; Alam, M.; Kobayashi, A. *J. Am. Chem. Soc.* **1976**, *98*, 1059-1060.

M. Wahid Khalil, Lois J. Durham, Carl Djerassi*

Department of Chemistry, Stanford University
 Stanford, California 94305

Donato Sica

Istituto di Chimica Organica
 via Mezzocannone 16, Napoli, Italy

Received November 21, 1979

Helixanes. The First Primary Helical Molecules: PolyoxapolySpiroalkanones

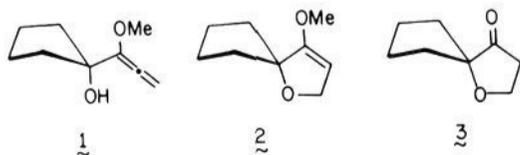
Sir,

The suggestion in the early fifties that the secondary structure of polypeptides has an α -helix conformation¹ led the way for extensive studies of helical topology, especially in biopolymers.² In particular the far-reaching double-stranded helical conformation for DNA³ and the helical nature of many polymers^{2,4} highlight the fact that helical molecules have a preminent place in macromolecular chemistry. Molecules that have helical topology but do not fall into the above classes are helicene,⁵ skewed paracyclophane,⁶ and helical triphenylmethane systems.⁷

All of the above helical molecules owe their helical topology to their secondary and tertiary structure.⁸ To date that is no molecule that is helical because of the primary bonding structure.

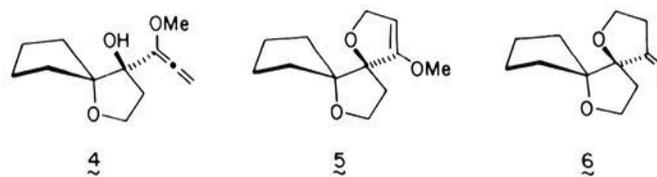
Here we report a rational synthesis of the *first primary helical molecules*, based upon the shape (bond angles and bond lengths) of the *tetrahydrofuran ring system*.

The adduct **1** between cyclopentanone (a starting block) and α -lithio- α -methoxyallene, on treatment with potassium *tert*-butoxide (0.2 equiv) in *tert*-butyl alcohol containing 18-crown-6 (0.05 equiv) heated at reflux for 15 h gave **2**.⁹ Acid hydrolysis (6 N H₂SO₄) of **2** gave **3**. The choice of conditions for the release of the dihydrofuranone carbonyl group is crucial in this step and in the subsequent hydrolysis procedures of the enol ethers described, since 1 N H₂SO₄ or two-phase systems (oxalic acid-dichloromethane) gave **3** contaminated with decomposition products, whereas increasing the strength of the acid to 6 N H₂SO₄ eliminated degradation and produced only a clean high-yield conversion of **2** into **3** (82% overall yield on a 8.0-g scale).

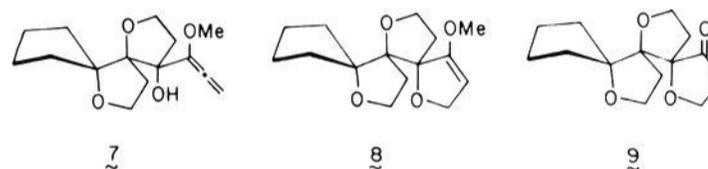


Since we started with a carbonyl compound, cyclopentanone, and spiroannulated a dihydrofuranone onto it, and since we have ended up with a new carbonyl compound, namely 1-oxaspiro[4.4]nonan-4-one (**3**), the product in principle is capable of being subjected to another spiroannulation sequence. The carbonyl group of **3** is relatively hindered, but it reacts cleanly with α -lithio- α -methoxyallene to give the adduct **4**, after aqueous ammonium chloride workup, in 95-98% yield.¹⁰ The adduct **4** on treatment with KOBu^t (0.1 equiv)/

HOBu^t/18-crown-6 (0.05 equiv) heated at reflux for 15 h gave **5**, which on acid hydrolysis (6 N H₂SO₄) gave **6** (70%).¹¹

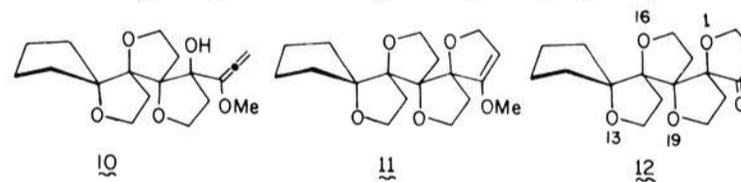


At this point we have reached a crucial stage. The first and only stereochemical issue arises. The carbonyl group in **6** has two diastereotopic faces available for the addition of a nucleophilic species. Dreiding models and space-filling models indicate that one face of **6** is severely hindered by the two five-membered rings already present. In particular the cyclopentane ring sits under one face of the carbonyl group in **6**. In the event, **6** adds α -lithio- α -methoxyallene to give a single crystalline adduct **7**, mp 133-137 °C in $\geq 90\%$ yield.¹² Treatment of this adduct with KOBu^t (0.2 equiv)/HOBu^t 18-crown-6 (0.05 equiv) heated at reflux led to **8**, which on acid hydrolysis (6 N H₂SO₄) gave the beautifully crystalline cyclopentyl[3]helixane **9**, mp 88.5-90 °C (67% overall from **7**).

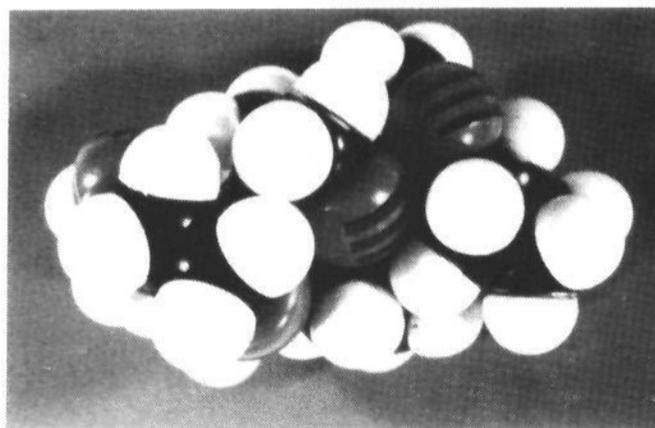


We hoped that, as we extended the spiro rings, the allenyl adducts would undergo ring closure under increasingly mild conditions. This optimistic view is based upon the idea that the steric compression of the alkoxide of **10** would favor cyclization since covalent association of a metal counterion (Li or K in the cases described) is sterically hindered, and furthermore **10** may act as its own crown ether ionophore and effectively remove the counterion (Li or K) from covalent association with the alkoxide.

Treatment of **9** with α -lithio- α -methoxyallene gave **10**, mp 92-99 °C (45%),¹³ which cyclized to **11** at room temperature (28 °C) when treated with KOBu^t (0.2 equiv)/HOBu^t/18-crown-6 (0.05 equiv) to give cyclopentyl[4]helixane **12**, mp 123-124 °C (67%), after acid (6 N H₂SO₄) hydrolysis. Models



(CPK) (see photograph) show that cyclopentyl[4]helixane **12** is a helix-shaped molecule. The oxygen atoms and methylene groups spiral around the central core of the molecule. To establish the structure of **12** unequivocally and confirm specu-



lations concerning stereochemistry, a single-crystal X-ray analysis was conducted. The results are as follows.

Cyclopentyl[4]helixane crystallized in the triclinic space group *P*1 with four molecules per unit cell. Accurate lattice parameters are $a = 13.881$ (4), $b = 8.853$ (2), $c = 13.668$ (3) Å; $\alpha = 104.83$ (2), $\beta = 76.60$ (2), $\gamma = 78.71^\circ$. The structure was solved by direct methods and has currently been refined